# Quick Manual



### ß Gal-Juice/ß Gal-Juice PLUS (1 033 11/ 1 033 12)

#### **Components included:**

β-Gal-Juice/plus	<b>50ml</b> Chemiluminescent Substrate for measurement of ß-Galactosidase Enzyme. <b>Store at +4°C.</b>

Triggering Reagent 50ml Store at +4°C

#### **Preparation of Cell Lysates:**

For cell lysis, we suggest the follow buffers:

- Animal cells: 100 mM potassium phosphate (pH 7,8), 0,2% Triton X100
- Yeast cells : 0,5 M sodium phosphate (pH 7,1) 50 mM KCl, 5 mM MgSO<sub>4</sub>

#### Standard Protocol for Detection of β-Galactosidase in Microtiter Plate Luminometer

#### Equilibrate at room temperature for 30 minutes before use.

Best results for chemiluminescent detection of  $\beta$ -Galactosidase enzyme or reporter assays can be obtained from 30 minutes to 60 minutes incubation of  $\beta$ -Gal Juice with  $\beta$ -Galactosidase enzyme and read the plate or tube within 2 to 10 minutes after triggering the reaction mixture by accelerator or enhancer.

- Add 5 to 10  $\mu$ l of diluted  $\beta$ -Galactosidase enzyme or cell extract to microplate wells.
- Add 50 to 100  $\mu l$  of  $\beta$ -Gal Juice and incubate for 30 to 60 minutes at room temperature.
- Add 50 to 100  $\mu$ l of triggering reagent and shake it for 10 seconds.
- Place the microtiter plate in luminometer and start reading.
- Best results are obtained between 2 to 10 minutes.

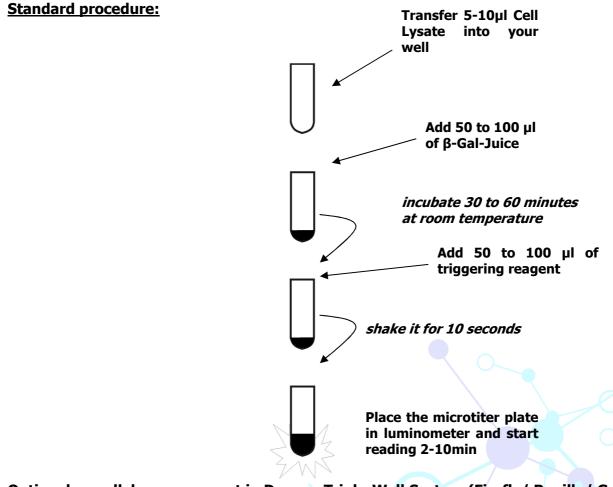
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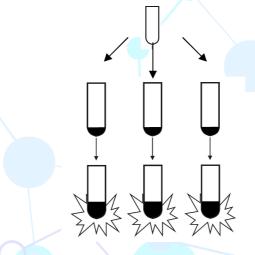


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#### <u>Optional: parallel measurement in Duo- or Triple-Well System (Firefly/ Renilla/ Gaussia/</u> <u>AP/ βGal)</u>

Split the cell lysate and transfer to different tubes for measuring several enzymes in one sample.



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